

Experimental and Molecular Medicine

Guide to Authors

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Instructions to Authors

All submitted manuscript should contain original research that has not been published previously and is not under consideration for publication elsewhere.

Manuscript submission

Manuscripts can be submitted online or by mail.

Online submission: For instructions, visit:

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Submission by mail: Submit **three** print copies of each manuscript including **one** original set with high quality graphics along with a Manuscript submission form (available at <http://www.e-emm.org/> or on page x). A letter must be included indicating the address, telephone and fax numbers, and E-mail address of the corresponding author. Send manuscripts and correspondence to:

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Submission of digital files: Following acceptance of a manuscript, authors should upload a digital file containing the final version of the manuscript via the online submission system. Microsoft Word is preferred, although other formats are generally acceptable. It is also encouraged to submit digital files of figures prepared in appropriate formats (see Tables and Figures). If

authors do not provide electronic files of their figures, they will be charged separately (about US\$5 for a simple line graph) and publication of the manuscript will be delayed. If the memory size of digital files exceeds the capacity of the online submission system, authors should submit them to EMM office via E-mail (ksbmb3@ksbmb.or.kr), or provide a CD or a USB by regular mail.

Manuscript and other materials will not be returned unless specifically requested.

Organization of the manuscript

Manuscripts must be typed or printed on 21×29.7 cm (A4 size) high-quality paper in **double spacing** throughout with at least 3 cm wide margins on all sides. The text must be typed in a font size of at least ten points. The manuscript is to be arranged in the following order: (a) title, author(s), and complete address(es) of institution(s); (b) abbreviations; (c) running title; (d) abstract and keywords; (e) introduction; (f) results; (g) discussion; (h) methods; (i) acknowledgements; (j) references; (k) figure legends; (l) tables; and (m) figures. Number all pages with the title page as page 1. To make papers more readable and informative, the EMM requests that authors mark the followings for typesetting in **italic**.

- (a) Biological name of organisms: *Saccharomyces cerevisiae*, *E. coli*
- (b) Restriction enzymes and some of enzymes: *EcoRI*, *Taq* polymerase
- (c) Name of genes: *src*, *c-H-ras*, *myc*
- (d) Latin: *in vivo*, *in vitro*, *in situ*
- (e) Some of chemical structure: *trans*-retinol, *cis*-acting, *N*-carbamoylaspartate
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Title: The title of the manuscript should be as short and informative as possible. It should not contain nonstandard abbreviations, subtitles, or colons, nor exceed two printed lines (about 18 words). The EMM reserves the right to reword titles, with the final approval of the authors. The title page should also give the names of all authors and their complete mailing addresses. The title page should also include the name, the telephone and fax numbers, and the E-mail address of the author to whom all correspondence about the manuscript, including proofs, will be sent.

Abbreviations: Standard abbreviations may be used without definition. Any nonstandard abbreviations should be spelled out on first use, followed by the abbreviated form in parentheses. Thereafter, abbreviated form may be used throughout the manuscript. Undefined abbreviations cannot be used. Provide a list of all nonstandard abbreviations in alphabetical order on a page following the title page.

Running title: The running title to be printed at the top of each page of a published paper cannot exceed 6 words.

Abstract: The abstract should succinctly and clearly describe the major findings reported in the manuscript. It must not exceed 250 words nor contain nonstandard or undefined abbreviations or specialized terms. It should be understandable in itself, since it is frequently used as an abstract in indexes.

Keywords: Six keywords should be appended to the abstract in alphabetical order. The keywords should be standard MeSH-Medline terms (<http://www.nlm.nih.gov/mesh>). The list submitted may be amended to ensure that entries are MeSH-Medline

terms.

Introduction: The introduction should present the purpose of the studies reported and their relationship to earlier work in the field. It should not be an extensive review of the literature nor, in general, exceed two typed pages.

Results: The results of experiments should be presented in figures and tables, although some results that do not require documentation can be given solely in the text. Extensive discussion should not be given in the Results section.

Discussion: The Discussion should be concise (usually less than four typed pages) and focused on the interpretation of the results rather than a repetition of the Results section.

Notes added in proof can be added to a manuscript only with the consent of the Editor.

Errors in a published paper should be corrected in the EMM in "Additions and Corrections."

Methods: The Methods should be as brief as possible but sufficiently complete to permit a qualified reader to repeat the experiments reported. Only truly new procedures should be described in detail; previously published procedures should be cited in references. Modifications of previously published procedures need to be given in detail only when this is necessary to repeat the work.

Supplemental data: Supplemental data are provided to support and enhance scientific information. EMM encourages authors to include such things as the detailed methodology, sequence alignment, background datasets and microarray hybridization experiments. Authors should submit the supplemental material in electronic format together with the article and supply a concise and descriptive caption for each file. Supplemental data will be reviewed as a part of the article. After acceptance, supplemental files will be published online alongside the electronic version of article in EMM web site. No editing will be done to the supplemental data at the editorial office and all changes are the responsibility of the author.

References: The references must be cited by Harvard system (author and date) not the numbering system. References should be quoted in the text with the author's name and date in parentheses, and listed at the end of the paper in alphabetical order. Journal names are abbreviated according to PubMed (<http://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=journals>). Authors are fully responsible for the accuracy of the references. "Personal communication" or "unpublished data" should be written in the text in parenthesis but not listed in references. References for journals and books should be in the following styles:

Journal articles

Lee JW, Cheong JH, Lee YC, Na SY, Lee SK. Dissecting the molecular mechanism of nuclear receptor action: transcription coactivators and corepressors. *Exp Mol Med* 2000;32:53-60

In press

Sun H, Wolfe JH. Recent progress in lysosomal-mannosidase and its deficiency. *Exp Mol Med* 2001; In press

Complete books

Halliwell B, Gutteridge JMC. *Free Radicals in Biology and Medicine*, 3rd Ed, 1999, Oxford University Press, Oxford, UK

Articles in books

Baird A, Bohlen P. Fibroblast growth factors. In *Peptide Growth Factors and Their Receptors* (Sporn MB, Roberts AB, eds), 1990, 369-418, Springer-Verlag, New York, NY

Tables and Figures

The number of tables and figures used to present data essential to illustrate or prove a point should be kept to a minimum. Very complex or large figures should be submitted in camera ready format typed or drawn in single space. For example, amino acid or nucleic acid sequences should always be prepared for direct photographic reproduction to avoid errors.

Tables should have titles and sufficient experimental detail in a legend immediately following the title to be understandable without reference to the text. Each column in a table must have a heading, and abbreviations, when necessary, should be defined in the legend.

Figures should have titles and explanatory legends that contain sufficient detail to make the figure easily understood. All legends should be typed consecutively in a separate section of the manuscript. Figures should be submitted in **final size (printed 1:1)**. They may be printed in either single column (80 mm width) or double column (165 mm width) format. The single column format is preferred. The size of text in figures should be 8-10 points, except for single letter markers which may be 12 points. The use of sans serif font such as Helvetica is preferred for all figure text. Numbers, letters, and symbols used in multi-paneled figures must be consistent. Use standard abbreviations whenever possible. For a line graph, use standard symbols in the following order of preference: ○, ●, □,

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On submission of the manuscript, a complete set of figures on paper about the same size as the manuscript pages should be included with each copy of the manuscript. Indicate on the front or back of each figure its number and the Running title. Only one set of top quality figures as line drawings is needed for the printer; other copies may be photographic prints or photocopies, except for electron micrographs or halftone figures where good quality prints must be supplied with each copy of the manuscript. Color figures should be supplied as continuous-tone prints on photographic-quality paper. Original figures will not be returned to authors after publication unless specifically requested.

Digital images. Authors are encouraged to submit digital images along with the reproduction quality hard copies of each figure, when the manuscript is accepted for publication. Image files can be submitted via online submission system, E-mail, or regular mail. The following guidelines can be used to create and submit digital files. *Format:* The EMM recommends EPS or TIFF files. Other application files (Powerpoint, Microsoft Word, Sigmaplot, MacDrawPro, etc.) are acceptable only when the resolution of images meets the publication quality. *Resolution:* 1,200 dpi bitmap mode for black and white images such as line graphs, 600 dpi grayscale mode for images such as gels or blots, and 600 dpi CMYK mode for color images.

EMM standard abbreviations

- A**, absorbance
A, ampere
Å, angstrom (10^{-10} m)
Ab, antibody
Ag, antigen
AIDS, acquired immunodeficiency syndrome
ANOVA, analysis of variance
AP-1, activator protein-1
APC, antigen-presenting cell
ATP, adenosine triphosphate (also **ADP**, **AMP**, **CTP**, **UDP**, etc.)
ATPase, adenosine triphosphatase (also **ADPase**, **AMPase**, etc.)
- bp**, base pair
Bq, Becquerel
BSA, bovine serum albumin
- $^{\circ}\text{C}$, degrees Celsius
cal, calorie
cAMP, cyclic AMP (also cGMP)
cDNA, complementary DNA
CFU, colony-forming unit
CHAPS, 3-[(3-cholamidopropyl)-dimethylammonio]-1-propanesulfonic acid
Ci, curie
CNS, central nervous system
CoA, coenzyme A
Con A, concanavalin A
COX, cyclooxygenase
cpm, counts per minute
CSF, colony-stimulating factor
CTL, cytotoxic T lymphocyte
- d**, day
d, density
Da, dalton; **kDa**, kilodalton
2D, 2-dimensional
DEAE, diethylaminoethyl
DMEM, Dulbecco's modified Eagle's medium
DMSO, dimethylsulfoxide
DNA, deoxyribonucleic acid
DNase, deoxyribonuclease
dpm, disintegration per minute
dsDNA, double-stranded DNA (also **dsRNA**)
DTT, dithiothreitol
- ED₅₀**, 50% effective dose
EDTA, ethylenediaminetetraacetic acid
EGF, epidermal growth factor; **EGFR**, epidermal growth factor receptor
EGFP, enhanced GFP
EGTA, ethylene glycol-bis(b-aminoethylether)-*N,N,N',N'*-tetraacetic acid
ELISA, enzyme-linked immunosorbent assay
ERK, extracellular signal-regulated protein kinase
- FACS**, registered trademark of Becton Dickinson and Company for a fluorescence-activated cell sorter
FAD, flavin-adenine dinucleotide
- FBS**, fetal bovine serum
Fc, crystalizable fragment of Ig molecule
FCS, fetal calf serum
FGF, fibroblast growth factor (**bFGF**, basic FGF)
FISH, fluorescence in situ hybridization
FITC, fluorescein isothiocyanate
FMN, flavin mononucleotide
Fox, Forkhead box
- g**, gram
g, unit of gravity
GAPDH, glyceraldehyde 3-phosphate dehydrogenase
G-CSF, granulocyte colony-stimulating factor
GFP, green fluorescent protein
GM-CSF, granulocyte/macrophage colony-stimulating factor
GSH, reduced glutathione; GSSG, oxidized glutathione
Gy, gray
- h**, hour
HA, hemagglutinin
HBSS, Hank's balanced salt solution
HBV, hepatitis B virus
HCV, hepatitis C virus
HDL, high-density lipoprotein
Hepes, *N*-2-hydroxyethylpiperazine-*N'*-2-ethanesulfonic acid
H&E, hematoxylin and eosin
HGF, hepatocyte growth factor
HIV, human immunodeficiency virus
HLA, human histocompatibility leukocyte antigens
HPLC, high-performance liquid chromatography
HRP, horseradish peroxidase
HUVEC, human umbilical vein endothelial cell
Hz, cycles per second
- ID₅₀**, 50% infective or inhibiting dose
IFN, interferon (e.g., IFN- γ)
Ig, immunoglobulin (IgG, IgE etc.)
IGF, insulin-like growth factor
IkB, inhibitor of κB
IL, interleukin (e.g., IL-2)
i.m., intramuscular
i.n., intranasal
i.p., intraperitoneal
IU, international unit
i.v., intravenous
- J**, joule
JAK, Janus kinase
JNK, c-jun N-terminal kinase
- k*, rate constant
K, degree absolute
kb, kilobase
K_d, dissociation constant
K_i, inhibition constant
KLH, keyhole limpet hemocyanin
K_m, Michaelis constant
KO, knockout

l, liter; ml, milliliter; μ l, microliter

LD₅₀, 50% lethal dose

LDL, low-density lipoprotein

LPS, lipopolysaccharide

LTR, long terminal repeat

m, meter

M, molar

mAb, monoclonal antibody

MALDI, matrix-assisted laser desorption/ionization

MAPK, mitogen activated protein kinase

Mb, megabase

2-ME, 2-mercaptoethanol

MEK, MAPK kinase

MEM, minimal essential medium

MHC, major histocompatibility complex

min, minute

MMP, matrix metalloproteinase

mo, month

mol, mole

Mops, 3-(*N*-morpholino)propanesulfonic acid

M_r, relative molecular mass

MRI, magnetic resonance imaging

MS, mass spectrometry

mTOR, mammalian target of rapamycin

MTT, 3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide

MW, molecular weight

n, number in study or group

NAD⁺, oxidized nicotinamide adenine dinucleotide (not NAD); also **NADP⁺**

NADH, reduced nicotinamide adenine dinucleotide; also

NADPH

ND, not determined

NF- κ B, nuclear factor kappa B

NK cell, natural killer cell

NKT, natural killer T (cell)

NMDA, N-methyl-d-aspartate

NMR, nuclear magnetic resonance

NO, nitric oxide

no., number

NOD, nonobese diabetic

NOS, nitric oxide synthase; **iNOS**, inducible NOS; **eNOS**, endothelial NOS

NP-40, Nonidet P-40

NSAID, nonsteroidal antiinflammatory drug

OD, optical density

ORF, open reading frame

OVA, ovalbumin

P, probability

PAGE, polyacrylamide gel electrophoresis

PBL, peripheral blood leukocyte

PBMC, peripheral blood mononuclear cell

PBS, phosphate-buffered saline

PCR, polymerase chain reaction

PDGF, platelet-derived growth factor; **PDGFR**, PDGF receptor

PFU, plaque-forming unit

pg, picogram(s)

PG, prostaglandin (e.g., PGE₂)

pH, hydrogen ion concentration

PHA, phytohemagglutinin

pI, isoelectric point

P_i, orthophosphate; **PP_i**, pyrophosphate

PI3K, phosphatidylinositol-3'-kinase

Pipes, piperazine-*N,N'*-bis(2-ethanesulfonic acid)

PKC, protein kinase C; **PKA**, protein kinase A;

PKB, protein kinase B

PLC, phospholipase C; **PLA**, phospholipase A;

PLD, phospholipase D

PMA, phorbol myristate acetate

PMSF, phenylmethylsulfonyl fluoride

PVDF, polyvinylidene difluoride

r, recombinant (e.g., rIFN- γ)

r, correlation coefficient

rad, radiation-absorbed dose

RANK, receptor activator of NF- κ B; **RANKL**, RANK ligand

RFLP, restriction fragment length polymorphism

RIA, radioimmunoassay

RLU, relative light unit(s)

RNA, ribonucleic acid; **mRNA**, messenger RNA; **tRNA**, transfer

RNA; **rRNA**, ribosomal RNA; **shRNA**, short hairpin RNA; **siRNA**, small interfering RNA

RNase, ribonuclease

rpm, revolution per minute

RPMI, Roswell Park Memorial Institute medium

RT-PCR, reverse transcription polymerase chain reaction

s, second

s, sedimentation coefficient

S, Svedberg unit of sedimentation coefficient

s.c., subcutaneous

SCID, severe combined immunodeficiency disease

SD, standard deviation

SDS, sodium dodecyl sulfate

SE, standard error

SEM, standard error of the mean

SH2, Src homology 2

SLE, systemic lupus erythematosus

SOCS, suppressor of cytokine signaling

SOD, superoxide dismutase

S-S, disulfide group

SSC, standard saline citrate

STAT, signal transducer and activator of transcription

SV40, simian virus 40

t_{1/2}, half-life, half-time

TBS, Tris-buffered saline

TCA, trichloroacetic acid

TCR, T cell receptor

TFA, trifluoroacetic acid

Tg, transgene, transgenic

TGF, transforming growth factor

Th, T helper cell (Th1 cell, etc.)

TLC, thin-layer chromatography

TNF, tumor necrosis factor

TOF, time-of-flight (MALDI/TOF)

Treg, regulatory T cell

Tris, tris(hydroxymethyl)aminomethane

TUNEL, terminal deoxynucleotidyl transferase-mediated dUTP nick end-labeling

TX, thromboxane (e.g., TXA)

U, unit

UV, ultraviolet

V, volt

VCAM, vascular cell adhesion molecule

VEGF, vascular endothelial growth factor; **VEGFR**, VEGF receptor

VLDL, very-low-density lipoprotein

V_{max} , maximum velocity

vol, volume

W, watt

wk, week

wt, weight

WT, wild-type

yr, year